statically by these anionic amphipathic activators. Such a factor was removed by partial purification on Sephadex column.

The authors greatly appreciate the kindness of Professor Dr. D. J. Hanahan, who showed us the Thesis of Miss M. E. Granade prior to publication, and also wish to thank Professor Dr. L. L. M. Van Deenen for generously supplying the synthetic lecithin.

Department of Medical Chemistry, Kansai Medical School, Moriguchi, Osaka (Japan) KUNIHIKO SAITO KIYOMI SATO

```
D. Fairbairn, J. Biol. Chem., 173 (1948) 705.
R. M. C. Dawson, Biochem. J., 68 (1958) 352.
R. M. C. Dawson, Biochem. J., 70 (1958) 559.
A. D. Bangham and R. M. C. Dawson, Biochem. J., 72 (1959) 486.
A. D. Bangham and R. M. C. Dawson, Biochem. J., 75 (1960) 133.
M. Kates, J. R. Madeley and J. L. Beare, Biochim. Biophys. Acta, 106 (1965) 630.
J. L. Beare and M. Kates, Can. J. Biochem. 45 (1967) 101.
D. N. Rhodes and C. H. Lea, Biochem. J., 65 (1957) 526.
D. J. Hanahan, J. C. Dittmer and E. Warashina, J. Biol. Chem., 228 (1960) 685.
K. Saito and D. J. Hanahan, Biochemistry, I (1962) 521.
C. S. Hanes and F. A. Isherwood, Nature, 164 (1949) 1107.
E. Chargaff, C. Levene and C. Green, J. Biol. Chem., 175 (1948) 67.
```

Received November 6th, 1967

Biochim. Biophys. Acta, 151 (1968) 706-708

BBA 63293

## Fluoroacetyl-CoA as a substrate for malate synthase

The toxicity of fluoroacetate to warm-blooded animals has been traced to a lethal synthesis of fluorocitrate, a potent inhibitor of aconitase<sup>1</sup>. The  $K_m$  for fluoroacetyl-CoA in the reaction with oxaloacetate catalyzed by citrate synthase was found<sup>2</sup> to be  $2.5 \cdot 10^{-5}$  M, the same as that for acetyl-CoA in this reaction.

Acetyl-CoA is a substrate for an analogous reaction catalyzed by malate synthase (L-malate glyoxylate-lyase (CoA-acetylating), EC 4.1.3.2), in which glyoxylate is the acetyl acceptor. DIXON, KORNBERG AND LUND³ in examining the substrate specificity of this enzyme, observed that fluoroacetyl-CoA was cleaved at a rate "approximately one-quarter of the rate observed with similar concentrations of acetyl-CoA".

We report here the results of experiments in which the kinetics of the reaction with fluoroacetyl-CoA are compared to those with acetyl-CoA as substrate. The malate synthase employed was that present in glyoxysomes prepared from the endosperm tissue of germinating castor beans<sup>4</sup>, with a specific activity of 0.6–1.4  $\mu$ moles/min per mg protein.

Enzyme activity was measured by following the rate of appearance of SH resulting from the cleavage of acetyl-CoA in the presence of glyoxylate<sup>5</sup>, with correction, as necessary, for the rate observed before addition of glyoxylate. In the ab-

Biochim. Biophys. Acta, 151 (1968) 708-710

sence of glyoxylate, acetyl-CoA was not cleaved by the enzyme at a measurable rate, and, as shown in Table I the rate observed when glyoxylate was present was proportional to the amount of malate synthase provided. When fluoroacetyl-CoA alone was added to the enzyme there was a slow production of SH, but the addition of glyoxylate resulted in a 3–4-fold stimulation. The corrected rate of reaction with

## TABLE I

RATES OF REACTION OF ACETYL-COA AND FLUOROACETYL-COA IN PRESENCE OF MALATE SYNTHASE AND GLYOXYLATE

The reaction mixtures contained, in a final volume of 1.0 ml: 50  $\mu$ moles potassium phosphate (pH 8.0), 7.5  $\mu$ moles MgCl<sub>2</sub>, I  $\mu$ mole 5,5'-dithiobis-2-nitrobenzoic acid (DTNB) (Aldrich Chemical Co.), 0.2  $\mu$ mole acetyl-CoA (ref. 6) or fluoroacetyl-CoA (refs. 7, 8) and enzyme as indicated. The change in  $A_{412~m\mu}$  was recorded for 3 min before adding I  $\mu$ mole sodium glyoxylate, and from the subsequent trace, initial rates of reaction were calculated. Temp.: 30°.

| Amount<br>of enzyme<br>added<br>(µg) | Sulfhydryl produced ( $\mu$ moles min $\times$ 100) in presence of |                       |  |  |
|--------------------------------------|--|-----------------------|--|--|
|                                      | Acetyl-<br>CoA +<br>glyoxylate                                     | Fluoro-<br>acetyl-CoA | Fluoro-<br>acetyl-CoA<br>+ glyoxylate* |  |
| 5                                    | 0,6  | _                     |  |  |
| 14                                   | 1.7  | _                     |  |  |
| 24                                   | 2.4  |                       | 0.35                                   |  |
| 38                                   | 4.0  | 0.15                  | 0.45                                   |  |
| 48                                   | 5.8  | 0.20                  | 0.75                                   |  |
| 96                                   | _  | 0.35                  | 1.35                                   |  |
| 144                                  |  | 0.55                  | 2.10                                   |  |

<sup>\*</sup> Corrected for fluoroacetyl-CoA.

fluoroacetyl-CoA increased with increasing enzyme concentration (Table I) and remained at roughly one-eighth of that observed with acetyl-CoA.

Fig. 1 shows the results obtained when the concentration of fluoroacetyl-CoA was varied in the presence of saturating glyoxylate. The  $K_m$  derived from these data is 1.1·10<sup>-5</sup> M.  $K_m$  values determined similarly for glyoxylate in the presence of an excess of fluoroacetyl-CoA and for the substrates of the normal malate synthase reaction

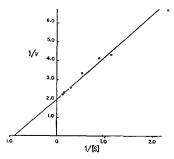


Fig. 1. Reciprocal plot of the effect of fluoroacetyl-CoA concentration S, (10<sup>-5</sup> M) on reaction velocity v (m $\mu$ moles consumed per min). Assay conditions as in Table I, with 60  $\mu$ g protein/ml and 3.3  $\mu$ moles glyoxylate/ml.

TABLE II  $K_m$  values for reaction of acetyl-CoA and fluoroacetyl-CoA with malate synthase

| $K_m$ values $(M)$ for |                          |  |
|------------------------|--------------------------|--|
| Acetyl-CoA             | Fluoro-<br>acetyl-CoA    | Glyoxylate                                   |
| 1.7 · 10-5             | <br>I.I·IO <sup>-5</sup> | 5.8·10 <sup>-5</sup><br>5.5·10 <sup>-5</sup> |
|                        | Acetyl-CoA               | Acetyl-CoA Fluoro-acetyl-CoA                 |

are given in Table II. The values obtained for this latter reaction are close to those observed previously for the enzyme from castor bean endosperm<sup>9</sup> and from other cells<sup>3</sup>. The  $K_m$  for glyoxylate when fluoroacetyl-CoA was the co-substrate is virtually the same as that observed in the presence of acetyl-CoA; that for fluoroacetyl-CoA is slightly lower than that for acetyl-CoA.

This work was supported by AEC Contract AT-11-1-330. G. L. P. was a Public Health Service Trainee (Grant 5T1 GM 1195-03).

Departments of Chemistry and Biological Sciences, Purdue University,

GARY L. POWELL\* HARRY BEEVERS

Lafayette, Ind. (U.S.A.)

- 1 R. A. Peters, Biochemical Lesions and Lethal Synthesis, MacMillan, New York, 1963, p. 88.
- 2 D. W. FANSHIER, L. K. GOTTWALD AND E. KUN, J. Biol. Chem., 239 (1964) 425.
- 3 G. H. DIXON, H. L. KORNBERG AND P. LUND, Biochim. Biophys. Acta, 41 (1960) 217.
- 4 R. W. Breidenbach and H. Beevers, Biochem. Biophys. Res. Commun., 27 (1967) 462.
- 5 B. Hock and H. Beevers, Z. Pflanzenphysiol., 55 (1966) 405.
- 6 E. R. STADTMAN, in S. P. COLOWICK AND N. O. KAPLAN, Methods in Enzymology, Vol. III, Academic Press, New York, 1957, p. 931.
- 7 R. O. Brady, J. Biol. Chem., 217 (1955) 213.
- 8 A. MARCUS AND W. B. ELLIOTT, J. Biol. Chem., 218 (1956) 823.
- 9 Y. YAMAMOTO AND H. BEEVERS, Biochim. Biophys. Acta, 48 (1961) 20.

## Received November 21st, 1967

Biochim. Biophys. Acta, 151 (1968) 708-710

<sup>\*</sup> Present address: Department of Biological Chemistry, School of Medicine, Washington University, St. Louis, Miss. 63110, U.S.A.